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## Expression and Characterisation of the Heavy Chain of Tetanus Toxin: Reconstitution of the Fully-Recombinant Dichain Protein in Active Form<sup>1</sup>

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Tetanus toxin, composed of a disulphide-linked heavy (HC) and light (LC) chain, preferentially blocks the release of inhibitory neurotransmitters in the spinal cord by Zn<sup>2+</sup>-dependent proteolytic cleavage of synaptobrevin. This intoxication involves binding via HC to ecto-acceptors on peripheral nerve endings, followed by internalisation and retrograde transportation to its prime site of action in central neurons. To facilitate exploitation of the toxin's unique activities, HC was expressed at a high level in *Escherichia coli* as a fusion with maltose binding protein; after cleavage by thrombin, free HC was isolated and its identity confirmed by Western blotting and N-terminal microsequencing. The expressed and native HC gave very similar circular dichroism spectra, excluding any gross differences in their folded structures. Recombinant HC antagonised the neuromuscular paralytic activity of the native toxin, by competing for binding to neuronal ecto-acceptors. The HC was reconstituted with bacterially-expressed LC to create disulphide-bridged dichain toxin that blocked neuromuscular transmission. The fully-recombinant toxin produced spastic paralysis in mice characteristic of the blockade of central inhibitory synapses, revealing that it undergoes axonal transport to the spinal cord, like the native toxin but with a reduced efficacy. This first report of the large-scale production of recombinant tetanus toxin in active form should facilitate studies on the use of engineered innocuous forms of the toxin as neuronal transport vehicles.

**Key words:** ecto-acceptors, expression, heavy chain, reconstitution, tetanus toxin.

Tetanus toxin (TeTx), produced by the strictly anaerobic, spore-forming rod-shaped bacillus *Clostridium tetani*, causes spastic paralysis by blocking neurotransmitter release at inhibitory synapses (1). This protein is synthesised as a single chain (~150 kDa) which is later cleaved by bacterial proteases to produce a dichain species, composed of a heavy (HC; ~100 kDa) and a light (LC; ~50 kDa) chain that are held together by strong non-covalent interactions and an inter-chain disulphide bridge (2). Although TeTx preferentially blocks the release of inhibitory transmitters in the central nervous system (CNS), at high

concentration it acts at the neuromuscular junction (NMJ) preventing the release of acetylcholine and causing flaccid paralysis (3-6). The action of TeTx involves a sequence of several steps (7): specific binding to neuronal ecto-acceptors, internalisation, translocation to the cytosol and, finally, inhibition of exocytosis due to cleavage of synaptobrevin by the Zn<sup>2+</sup>-dependent protease activity of its LC (8, 9) and, to some extent, by a protease-independent activation of neuronal transglutaminase(s) (10, 11).

The two constituent chains of TeTx can be separated with retention of their biological activities; also, when reconstituted they form the disulphide linked dichain toxin (2). Treatment of the isolated HC with papain (12) generates H<sub>C</sub> (~50 kD; C-terminal half of HC) and H<sub>N</sub> (~50 kD; N-terminal half of HC); these two fragments and LC appear to represent functional domains that are concerned with the multi-phasic intoxication process. HC or H<sub>C</sub> can compete with TeTx for binding to its acceptors on cultured cells (13-15) and antagonise the neuromuscular paralytic action of TeTx (13, 14). Also, H<sub>C</sub> is known to bind to certain gangliosides on the surface of target cells (15, 16) and undergo retrograde axonal transport, albeit to a limited extent (17), whilst H<sub>N</sub> has been shown to form channels in cell membranes (18) and contribute to the translocation of TeTx across membranes (19). LC is responsible for the intracellular blockade of transmitter release (8, 9, 20, 21).

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Abbreviations: TeTx, tetanus toxin; LC and HC, light and heavy chains of TeTx; H<sub>C</sub> and H<sub>N</sub>, C- and N-terminal halves of HC; LD<sub>50</sub>, amount of toxin that kills half of the injected mice within 4 days; factor Xa, activated factor X protease; DTT, dithiothreitol; SDS-PAGE, sodium dodecyl sulfate-polyacrylamide gel electrophoresis; MBP, maltose binding protein; MCS, multiple cloning site; PCR, polymerase chain reaction; IPTG, isopropyl-β-D-thiogalactoside; PMSF, phenylmethanesulfonyl fluoride; LB, Luria-Bertani medium; LA, Luria-Bertani agar; CNS, central nervous system; NMJ, neuromuscular junction; BSA, bovine serum albumen; KR and mKR, normal and modified Krebs Ringer solution.

After the initial cloning of TeTx gene (22), H<sub>c</sub> was successfully expressed in *Escherichia coli* (23), yeast cells (24), and insect cultured cells (25). The recombinant H<sub>c</sub> (r-H<sub>c</sub>) was shown to bind to ganglioside (26) and to protect mice against a challenge with TeTx (27, 28). There have been numerous reports on the ability of H<sub>c</sub> to transport exogenous proteins into CNS (29–32). These demonstrated that large proteins, whose entry to CNS is limited by the blood-brain barrier, may gain access through endocytosis in the periphery when conjugated to H<sub>c</sub>. Moreover, attachment of H<sub>c</sub> to horseradish peroxidase seemed to enhance the stability of this protein (31), as well as promoting its movement by trans-synaptic transport. However, whole TeTx binds with far higher affinity to neurons than H<sub>c</sub> and is also much more efficient in undergoing retrograde axonal transport, under physiological conditions (2, 17), indicating that H<sub>n</sub> fragment and/or LC of the toxin may contribute indirectly to the binding and transport of TeTx.

Although active recombinant LC (r-LC) has been successfully produced in *E. coli* (33, 34), the preparation of r-HC has not been reported, even though its use instead of native HC is particularly important for reconstitution with

enzymically-inactive mutated LC (33) to form an innocuous dichain which has great potential as a novel transporter. Therefore, a fusion of HC with maltose binding protein (MBP) was expressed in *E. coli* and, after purification of free r-HC, its properties were compared with those of H<sub>c</sub> prepared in similar manner. As fully-recombinant TeTx would be useful for elucidating the mechanisms underlying its acceptor-mediated internalisation and retrograde transport, this was obtained, for the first time, by reconstitution of expressed LC with r-HC and the resultant toxin shown to block neuroexocytosis and induce spastic paralysis.

## MATERIALS AND METHODS

**Materials**—Restriction endonucleases, DNA modifying enzymes and DNA purification kits were from Promega and used as recommended by the manufacturer. pMAL-c2 vector containing a factor Xa site was purchased from New England Bio-Labs; pMAL-c2-T is a modified version of the latter produced by insertion of a histidine tag and a thrombin susceptible site (see later). Thrombin, factor Xa and Sepharose 6B were obtained from Sigma. Purified

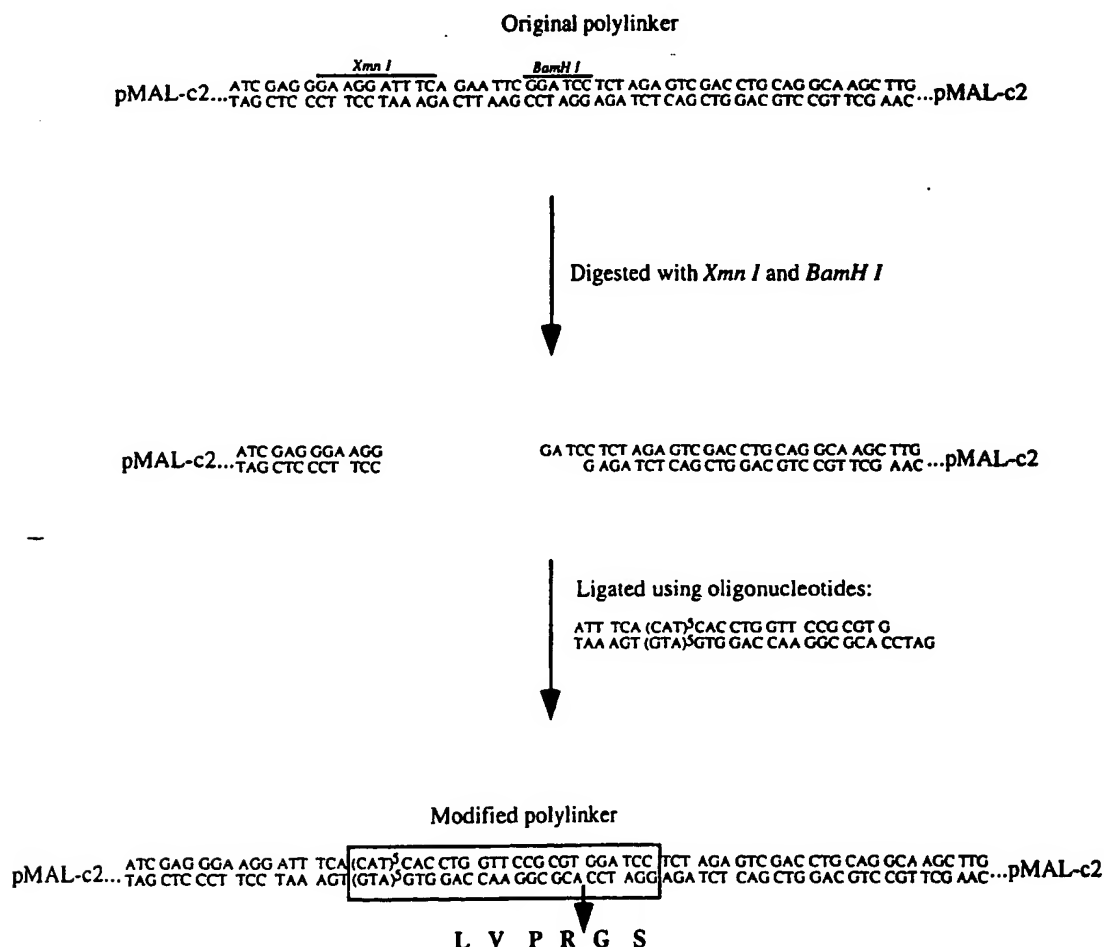


Fig. 1. Schematic representation of the construction of the expression vector pMAL-c2-T by insertion of a modified polylinker into pMAL-c2. The original polylinker was removed from pMAL-c2 by digestion with restriction-enzymes *XmnI* and *BamHI*. The resultant fragments were ligated using the two specified oligonucleotides to generate a modified polylinker (boxed), containing six

consecutive histidines [(CAT)<sup>5</sup>CAC] and a thrombin site. The product was subcloned into pMAL-c2, cut with the above-noted enzymes, and the sequence of the new vector pMAL-c2-T verified by DNA sequencing. The amino acid sequence recognised by thrombin is shown together with the peptide bond cleaved (↓).

TeTx, native LC, and HC were kindly provided by Dr. U. Weller.

**Preparation of pMAL-c2-T**—The vector pMAL-c2 has been widely used to express in *E. coli* foreign proteins fused with MBP via a factor Xa cleavage site in this laboratory (33, 35). As factor Xa is expensive and relatively inefficient in cleaving MBP from its fusion with HC of TeTx (unpublished observation), this vector was improved by introducing nucleotide sequences encoding a thrombin site and six consecutive histidines into the multiple cloning site (MCS) of pMAL-c2. These modifications were made as outlined in Fig. 1. Briefly, two oligonucleotides corresponding to the sense and antisense sequence of the polylinker were synthesised, phosphorylated and mixed together at 100°C for 5 min; annealing was promoted by cooling down the mixture slowly (less than 1°C/min) to 22°C. About 0.1 µg of the annealed polylinker was ligated with 10–20 ng of pMAL-c2 cut by *Xmn*I and *Bam*HI at 15°C for 16 h. The product was transformed into *E. coli* TG1 strain and spread onto Luria-Bertani agar (LA) plates containing 100 µg/ml of ampicillin; colonies containing the new polylinker in pMAL-c2 were confirmed by DNA sequencing of the modified junction in the isolated vector, using Sequenase version 2.0 DNA sequencing kit (USB).

**Construction of MBP-HC and MBP-H<sub>c</sub> Plasmids**—Oligonucleotides used as primers in the amplification of HC and H<sub>c</sub> genes were synthesised on a DNA/RNA synthesiser (Applied Biosystems). The whole HC gene was created by ligating its three fragments from pTet8, pTet14 and pTet215; these had been cloned into different vectors and were kindly provided by Dr. N.F. Fairweather. First, the

gene encoding H<sub>N</sub> was amplified by PCR using primers *a* (5'-AAT AGA TCT AGA TCA TTA ACA GAT TTA GGA-3') and *b* (5'-TTC TAA AGA TCT ATA CAT TTG ATA ACT-3') from pTet8; primers *c* (5'-ATG TAT AGA TCT TTA GAA TAT CAA GTA-3') and *d* (5'-ATC GAT AAG CTT TTA TCA GTC GAC CCA ACA ATC CAG ATT TTT AGA-3') from pTet14. The two PCR products were joined together by ligation via the *Bgl*III (AGA TCT) site, creating the H<sub>N</sub> gene product. H<sub>N</sub> DNA was cut by *Xba*I (TCT AGA) and *Hind*III (AAG CTT) and subcloned into pMAL-c2-T which had been cut with the same two enzymes, generating pMAL-H<sub>N</sub> plasmid. The whole HC gene was produced by joining the following fragments together: (i) H<sub>N</sub> released from pMAL-H<sub>N</sub> plasmid by *Sac*I (GAG CTC) and *Sal*I (GTC GAC); (ii) H<sub>c</sub> cut by *Sal*I and *Bam*HI from plasmid pTet215; and (iii) *Sac*I and *Bam*HI cut pMALc2-T vector, creating the pMAL-HC plasmid. For the preparation of pMAL-H<sub>c</sub> construct, primers *e* (5'-AAT AGA GGA TCC AAC GAA GAA GAC ATC-3') and *f* (5'-GCT AGC TCT AGA TTA TCA GTC GTT GGT-3') were employed to amplify its coding sequence from pTet215. The *Bam*HI (GGA TCC) and *Xba*I (TCT AGA) digested PCR product was subcloned into a pMAL-c2 cut with same pair of enzymes. The ligation mixture was incubated at 15°C for 16 h. Competent *E. coli* cells (JM109 strain for pMAL-HC and TG1 strain for pMAL-H<sub>c</sub>) prepared in ice-cold 75 mM CaCl<sub>2</sub> buffer were used for transformation. Recombinant clones were screened on LA plates containing 100 µg/ml of ampicillin and the identity of the junction in the DNA insert was confirmed by sequencing. The pMAL-HC possesses a sequence encoding six consecu-

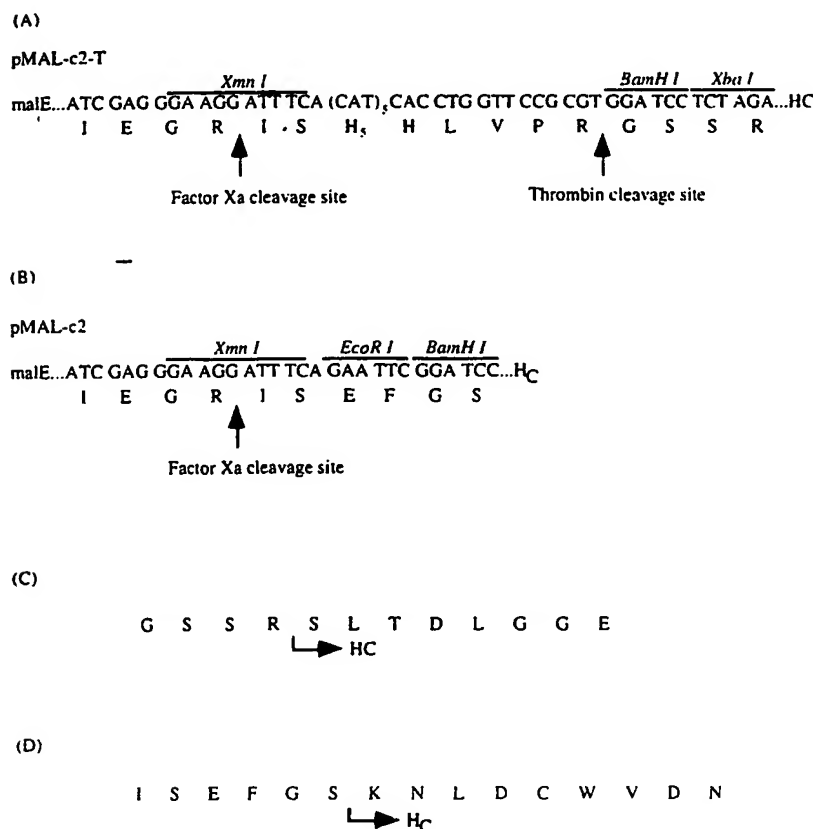


Fig. 2. Schematic presentation of pMAL-HC and pMAL-H<sub>c</sub> used for expressing r-HC and r-H<sub>c</sub>; verification of the N-terminal sequence of r-HC and r-H<sub>c</sub>. (A) Illustrates the MCS of pMAL-c2-T, including the factor Xa cleavage site, six consecutive histidines (His-tag) and thrombin cleavage site, which links MBP and HC. (B) Represents MCS of pMAL-c2, including the factor Xa cleavage site, between MBP and H<sub>c</sub>. (C) Shows the first 12 residues found at the N-terminus of r-HC protein purified from *E. coli*; microsequencing proved that the eight residues shown (arrow) at the N-terminus of r-HC are identical to those of native HC, preceded by the four amino acids encoded by the MCS sequence of pMAL-c2-T. (D) Represents the first 15 amino acids of the sequence obtained for the N-terminus of r-H<sub>c</sub>: nine of these amino acids (arrow) are the same as those in native H<sub>c</sub>, preceded by the six residues encoded by the MCS sequence of pMAL-c2.

tive histidines and cleavage sites for factor Xa and thrombin (Fig. 2A) whereas the pMAL-H<sub>c</sub> construct contains a factor Xa cleavage site sequence between MBP and H<sub>c</sub> protein (Fig. 2B).

**Preparation of Amylose Affinity Resin by Coupling of Starch to Sepharose 6B**—This was achieved using a modified protocol from Hermanson *et al.* (36). Sepharose 6B (100 ml) was washed with 2 liters of distilled water and then in 200 ml of 1 M Na<sub>2</sub>CO<sub>3</sub> (pH 11) before being resuspended in 40 ml of the latter buffer. Five milliliters of divinylsulfone was added and the suspension slowly mixed at 22°C for 60 min. The activated matrix was washed with 4 liters of distilled water and then suspended in 100 ml of 6% (w/v) soluble starch in 1 M Na<sub>2</sub>CO<sub>3</sub>. The mixture was continuously rotated at 50°C for 1 h before the addition of 5 ml of 2-mercaptoethanol to quench the unreacted divinylsulfone. After 30 min at 22°C, the resin was washed with 1 liter of column buffer [10 mM Tris-HCl (pH 7.4), 200 mM NaCl, 1 mM ethylene glycol-bis(β-aminoethyl ether) N,N,N',N'-tetraacetic acid, and 1 mM dithiothreitol (DTT)]. The resin was stored in 0.05% (w/v) sodium azide at 4°C before use.

**Expression and Purification of r-HC and r-H<sub>c</sub> Proteins**—Overnight cultures of bacteria containing either pMAL-HC or pMAL-H<sub>c</sub> construct was diluted (1:100) into fresh Luria-Bertani medium (LB) containing ampicillin (100 μg/ml), and grown for 2–2.5 h with shaking (200 rpm) at 37°C to reach the appropriate cell density ( $A_{600} \sim 0.5$ ). Isopropyl-β-D-thiogalactoside (IPTG) was added to a final concentration of 0.2 mM and the cultures shaken for an additional 3 h at 37°C (for r-H<sub>c</sub>) and 16–20 h at 30°C (for r-HC). The cells were harvested by centrifugation at 6,000 × *g* for 30 min at 4°C and resultant pellets were resuspended in the column buffer containing 1 mM phenylmethanesulfonyl fluoride (PMSF) before lysis by sonication for 10 min (10 s pulses plus an interval of 30 s) on setting 10 (maximal scale; Ultrasonic Processor, Model XL2020, Heat System, UK). After centrifugation of the cell lysate at 9,000 × *g* for 30 min at 4°C, the supernatants were loaded onto amylose affinity columns (2.5 × 10 cm, 40 ml resin) equilibrated with the column buffer. Following removal of unbound proteins by washing with the column buffer (320 ml), the bound MBP-HC or MBP-H<sub>c</sub> fusion protein was eluted with the same buffer containing 10 mM maltose. The fusion proteins were concentrated and cleaved either by thrombin (for MBP-HC) at 4°C for 16 h (1 unit to 75 μg of the fusion protein) or by factor Xa (for MBP-H<sub>c</sub>) at 22°C for 24 h [at an enzyme:protein ratio of 0.5–1:100 (w/w)]. The digestion was terminated by adding PMSF (final concentration 1 mM) to inactivate the excess proteases and the reaction mixture analysed by sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS-PAGE). The cleaved fusion proteins, after dialysis against the column buffer to remove the free maltose, were isolated from free MBP by readsorption of the latter to a new amylose column. The purified r-HC or r-H<sub>c</sub> were analysed by SDS-PAGE and Western blotting. N-terminal sequencing of r-HC and r-H<sub>c</sub> was carried out by subjecting the sample to SDS-PAGE, transfer onto polyvinylidene difluoride membranes (37) and automated Edman degradation using a model 4000 protein sequencer (Chelsea Instruments, London).

**CD Measurement of Native HC and r-HC**—Far-UV CD spectra were recorded on an Aviv 62DS, with an acceptance

angle ≥ 90°, fitted with a thermostated cell holder, using one 1 mm pathlength suprasil cuvette. Data points were collected in steps of 0.2 nm, from 300 to 190 nm. The CD spectra were recorded at 22°C at concentrations of native HC and r-HC at 0.10 mg/ml after dialysis against 10 mM sodium phosphate buffer (pH 7.0) at 4°C for 24 h. Each CD spectrum is the average of 5 scans from which the corresponding averaged baseline spectrum, obtained under identical conditions, has been subtracted. The results are expressed as mean residue ellipticity, taking a mean residue molecular mass for HC of 114.7, which was calculated from its amino acid sequence (38). Data were analysed to estimate the percentage of secondary structure, using a least squares algorithm (39) with reference data from Chang *et al.* (40).

**Measurement of the Effect of Native HC, r-HC, or r-H<sub>c</sub> on the Blockade by TeTx of Neuromuscular Transmission**—Mouse left phrenic nerve-hemidiaphragm, dissected from mice (T/O strain, 4 weeks old and ~20 g in weight), was immediately transferred into a closed circulatory superfusion system containing 10 ml of a modified Krebs-Ringer solution [mKR, consisting of: 118 mM NaCl, 4.7 mM KCl, 5 mM MgSO<sub>4</sub>, 0.5 mM CaCl<sub>2</sub>, 23.8 mM NaHCO<sub>3</sub>, 1.2 mM KH<sub>2</sub>PO<sub>4</sub>, 11.7 mM glucose, pH 7.4 (41, 42)] bubbled with 95% O<sub>2</sub> and 5% CO<sub>2</sub>, and supplemented with 0.1% (w/v) bovine serum albumen (BSA) to diminish non-specific adsorption of the toxins. The tissues were first incubated in mKR buffer alone (as a control) or the latter containing 500 nM native HC, 1 μM r-HC, 2 μM r-H<sub>c</sub>, or 2 μM MBP, at 4°C for 60 min; 1 nM TeTx was then added into the bath and co-incubated at 4°C for a further 30 min. To determine whether these proteins could displace bound TeTx, 1 nM TeTx was first incubated with the diaphragm in mKR buffer at 4°C for 30 min, followed by incubation without (as a control) or with 500 nM native HC, 1 μM r-HC, or 2 μM r-H<sub>c</sub> for a further 60 min. The diaphragms, after being washed thrice with mKR and with KR buffer (mKR with 2.5 mM CaCl<sub>2</sub> and 1.2 mM MgSO<sub>4</sub>) for another three times, were kept in a bath containing 10 ml KR buffer before the temperature was raised to 37°C. Muscle twitch was evoked by supra-maximal stimulation (Grass model S48 stimulator) of the phrenic nerve with bipolar electrodes and recorded via a force-displacement transducer (Electromed, UK) connected to an amplifier and computer system (MacLab, AD Instruments, UK). Parameters of nerve stimulation were 0.2 Hz square waves of 0.1 milliseconds duration with 1.5–2.5 V amplitude. Toxin-induced paralysis of neuromuscular transmission was calculated as the time taken for nerve-evoked muscle contraction to decrease to 10% of original value, as described previously (43).

**Reconstitution of Native and Recombinant Dichains and Assessment of Their Biological Activities**—LC of TeTx was expressed in *E. coli* as a fusion with MBP and purified on amylose resin; after cleavage with protease, LC was separated from MBP by affinity chromatography as detailed previously (33). Reconstitution was achieved by mixing equimolar amount of native (2) or recombinant HC and LC. The mixtures were dialysed against 50 mM Tris-HCl (pH 8.4), 1 M NaCl, 2 M urea, and 20 mM DTT with stirring at 4°C for 18 h, followed by dialysis against 50 mM Tris-HCl (pH 8.4) and 600 mM glycine without agitation at 4°C for another 72 h, as detailed previously (33).

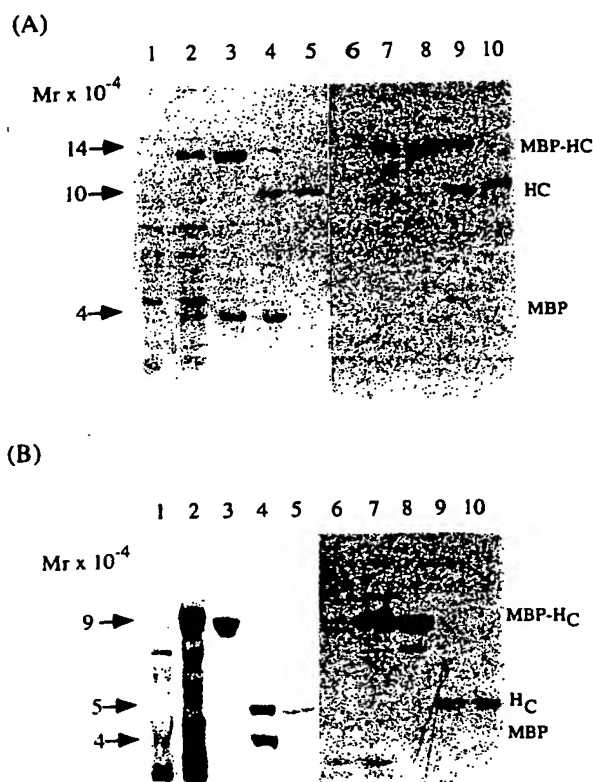
The extent of formation of dichain species was monitored by SDS-PAGE in the absence of reducing reagent. The relative neuromuscular activities of reconstituted preparations of recombinant and native dichain were assessed by measurement of neuromuscular paralysis *in vitro*, and determination of mouse lethality. For the former, mouse left phrenic hemi-diaphragm was dissected and maintained in the bath containing 10 ml KR buffer. After a stabilisation period of 10 min in the bath, each reconstituted sample or KR buffer (as a control) was added and nerve-evoked muscle contraction recorded as above. For quantitation of mouse lethality, toxin samples (maximum volume: 200  $\mu$ l/mouse) were injected subcutaneously into the dorsal neck region of mice (4-5 weeks old with 20 g body weight, four mice/group) and the LD<sub>50</sub> values (from three different experiments) determined, as described previously (41).

## RESULTS

**Characterisation of Plasmids Containing HC and H<sub>C</sub> Coding Sequences of TeTx**—The PCR amplified and purified H<sub>N</sub> and H<sub>C</sub> fragments of TeTx were digested with the appropriate restriction enzymes and subcloned into pMAL-c2-T and pMAL-c2, respectively. pMAL-HC was produced by ligating H<sub>N</sub> and H<sub>C</sub> into pMAL-c2-T, as detailed in "MATERIALS AND METHODS." DNA encoding HC or H<sub>C</sub> from positive colonies grown on LA plates containing ampicillin was purified using a DNA miniprep kit. DNA sequencing demonstrated that the two DNA fragments subcloned into pMAL vectors were in the correct reading frames with those for MCS, factor Xa and/or thrombin cleavage sites (Fig. 2, A and B). This creation herein of the modified pMAL-c2-T makes it possible to purify the HC fusion protein either on amylose resin *via* MBP (used in this study) or by chromatography on a nickel-containing column (*via* His-tag; data not shown); also, cleavage of MBP from the HC can be achieved with either factor Xa or thrombin. A major advantage of using thrombin is that efficient digestion can be accomplished at 4°C (see below) which minimises denaturation and maintains the solubility of HC which tends to become insoluble when incubated at higher temperatures for extended periods. In the case of MBP-H<sub>C</sub>, the unmodified pMAL-c2 construct (Fig. 2B) was used because digestion with factor Xa at 22°C proved satisfactory due to the greater solubility of H<sub>C</sub>.

**Expression and Purification of r-HC and r-H<sub>C</sub>**—The resultant plasmids (pMAL-HC and pMAL-H<sub>C</sub>) were transformed into *E. coli* JM109 and TGI, respectively, and induction of the fusion proteins was initiated by addition of IPTG. Affinity chromatography of the expressed protein was carried out as described previously (33) except that amylose resin was prepared rather than purchased; its binding capacity for the MBP-fusion proteins was similar to that of commercial resin. The expression and purification of MBP-HC and MBP-H<sub>C</sub> were monitored by SDS-PAGE; Coomassie staining showed that MBP-HC or MBP-H<sub>C</sub> appeared only in the lysates of cells induced by IPTG (Fig. 3, A and B). The bulk of each fusion protein became bound to the resin (not shown) and could be eluted from the amylose column with maltose (Fig. 3, A and B). Most of the MBP-HC was cleaved with thrombin after digestion at 4°C (Fig. 3A); at this low temperature the released HC remained soluble. Cleavage of MBP-H<sub>C</sub> required incubation with

factor Xa at 22°C (Fig. 3B). Free HC and H<sub>C</sub> were separated from MBP by reabsorption of the latter onto new amylose columns and their identities were confirmed by Western blotting, using mouse anti-serum raised against native HC (Fig. 3, A and B). Microsequencing revealed that eight residues at the N-terminus of r-HC are identical to those of native HC, preceded by the four amino acids encoded by the MCS nucleotide sequence of pMAL-c2-T (Fig. 2C); nine



**Fig. 3. Expression and purification of r-HC and r-H<sub>C</sub> monitored by SDS-PAGE and Western blotting.** *E. coli* was transformed with pMAL-HC or pMAL-H<sub>C</sub>; after induction with IPTG, the bacteria were pelleted, resuspended in column buffer and lysed by sonication. The cell lysate was loaded onto a column containing the amylose affinity resin (40 ml). After washing away unbound contaminants with 320 ml of column buffer, the bound MBP-HC or MBP-H<sub>C</sub> protein was eluted with 100 ml of column buffer (3 ml fractions) containing 10 mM maltose. The eluted fusion proteins (peak fractions) were concentrated in a Centricon (*M<sub>r</sub>* 50K cut-off). MBP-HC and MBP-H<sub>C</sub> were cleaved with thrombin at 4°C for 16 h, and factor Xa at 22°C for 24 h, respectively. After digestion, PMSF was added to a final concentration of 1 mM to inactivate the excess enzyme; MBP was removed by adsorption to a new lot of amylose resin. The samples were subjected to SDS-PAGE (8% polyacrylamide gel) with Coomassie staining (lanes 1-5) or detection with polyclonal anti-HC antibodies (lanes 6-10), by the procedure detailed elsewhere (33). Panels (A) and (B) show HC and H<sub>C</sub> preparation, respectively. Lanes 1 and 6: lysate of cells not induced; 2 and 7: lysate of cells induced with IPTG; 3 and 8: fusion protein eluted from the amylose column; 4 and 9: fusion protein after incubation with thrombin or factor Xa to release toxin sample and MBP; 5 and 10: purified toxin sample. The sizes of MBP-HC, MBP-H<sub>C</sub>, r-HC, r-H<sub>C</sub>, and MBP, derived from the mobilities of standard proteins, are shown. Note: A low molecular weight band shown in the uninduced and induced samples in (B) was probably caused by non-specific binding of anti-HC antibody to an unknown bacterial protein and after bio-specific elution from the amylose resin, the stained band disappeared.

residues obtained from the N-terminus of r-HC correspond to those in native H<sub>C</sub>, preceded by six amino acids encoded by the MCS nucleotide sequence of pMAL-c2 (Fig. 2D). After purification, ~15 mg MBP-HC and ~23 mg MBP-H<sub>C</sub> proteins were generally obtained from 1 liter of cell culture, respectively.

**CD Spectra of Native HC and r-HC**—The far UV CD spectrum of r-HC closely resembles that of its native counterpart (Fig. 4). The estimated secondary structure content for r-HC is 25%  $\alpha$ -helix, 37%  $\beta$ -sheet, 7%  $\beta$ -turn, and 31% random coil. HC isolated from TeTx using 4 M urea has been reported (44) to give somewhat different values; the lower proportion of  $\beta$ -structure may be attributed to the complete and irreversible loss of toxicity of TeTx known to be caused by 4 M urea (2), unlike the milder conditions employed herein. Notably, a similar content of  $\beta$ -structure (40%) has been published (4) for the homologous HC of botulinum neurotoxin type A. Our results for the r-HC are similar to those reported by de Filippis *et al.* (45) for the LC (27%  $\alpha$ -helix and 43%  $\beta$ -structure) of TeTx

and suggest that both chains may have similar secondary structures.

**Antagonism of the Neuromuscular Blocking Action of TeTx by Native HC, r-HC, or r-H<sub>C</sub>**—It has been shown that pre-incubation of diaphragm with native HC or H<sub>C</sub> derived from TeTx antagonises the neuromuscular blocking activity of TeTx by competing for acceptor sites on the nerve membrane (13, 14). This provided a means of assaying the biological activity of these recombinant proteins. Mouse phrenic nerve-muscle preparations were pre-equilibrated at 4°C for 60 min in mKR buffer alone (as a control) or with the sample specified, then incubated in the same buffer at

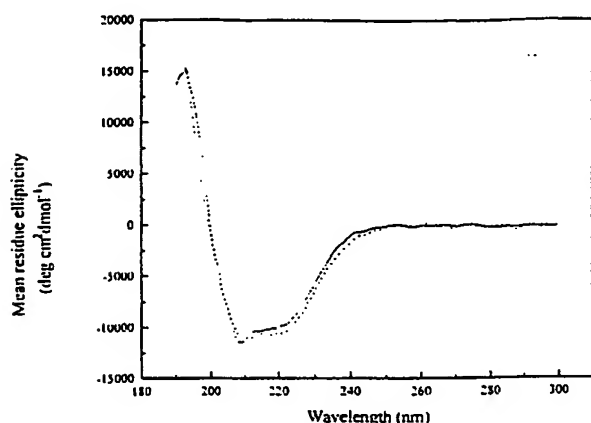


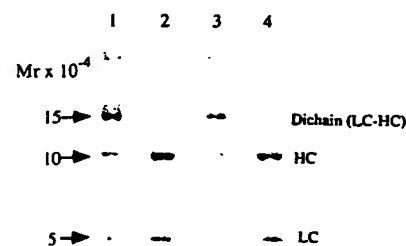
Fig. 4. Far UV CD spectra of r-HC and native HC. These spectra of HC were recorded on an Aviv 62DS instrument, with an acceptance angle  $\geq 90^\circ$ , fitted with a thermostatted cell holder, using a 1 mm pathlength suprasil cuvette. Data points were collected in steps of 0.2 nm, from 300 to 190 nm. UV CD spectra were recorded at 22°C for recombinant (—) and native (·····) HC at 0.10 mg/ml in 10 mM sodium phosphate buffer (pH 7.0).

TABLE I. Relative abilities of HC and H<sub>C</sub> to antagonise the blocking action of TeTx on neuromuscular transmission.

Proteins added <sup>a</sup> ( $\mu$ M; final conc.)	Paralysis time <sup>b</sup> (min)	<i>p</i> value <sup>c</sup>
None	250 $\pm$ 8	
Native HC (0.5)	369 $\pm$ 13	< .01
r-HC (1.0)	380 $\pm$ 9	< .01
r-H <sub>C</sub> (2.0)	376 $\pm$ 11	< .01
MBP (2.0)	246 $\pm$ 10	

<sup>a</sup>Isolated mouse nerve-muscle diaphragm preparations were used. Control tissue was incubated without the fragments at 4°C whereas the experimental groups were pre-incubated at 4°C with test proteins in mKR buffer for 60 min, and then exposed at 4°C to 1 nM TeTx for an additional 30 min. All tissues were washed thrice with mKR and three times with KR medium. The temperature was raised to 37°C, the phrenic nerve stimulated supramaximally and paralysis time recorded as detailed in Ref. 42. <sup>b</sup>Mean values ( $\pm$  SD) obtained from three different experiments. <sup>c</sup>Differences evaluated statistically using the Student's *t*-test.

(A)



(B)

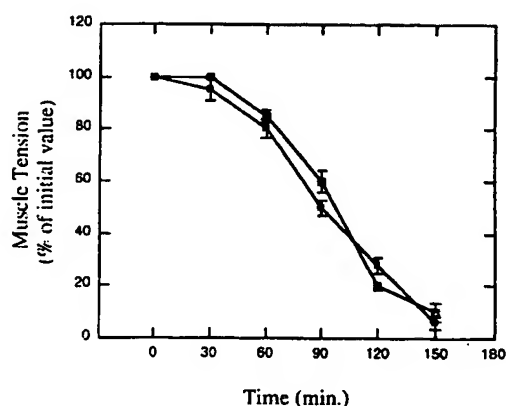


Fig. 5. Formation of disulphide-linked dichain toxin following reconstitution of either recombinant LC and HC or their native counterparts: effects on mouse neuromuscular transmission. Equimolar amounts of either recombinant LC and HC or their native equivalents were mixed together and dialysed against 50 mM Tris-HCl (pH 8.4), 1 M NaCl, 2 M urea, and 20 mM DTT with stirring at 4°C for 16 h, followed by dialysis against 50 mM Tris-HCl (pH 8.4) and 600 mM glycine without agitation at 4°C for 72 h, as detailed previously (33). (A) The formation of dichain was monitored by SDS-PAGE under non-reducing conditions. Lanes 1 and 2: dichain reconstituted from native LC and HC run under non-reducing (lane 1) and reducing (lane 2) conditions; lanes 3 and 4: dichain assembled from recombinant LC and HC run under non-reducing (lane 3) and reducing (lane 4) conditions. *M<sub>r</sub>* values for dichain, HC and LC, determined from their mobilities relative to standard proteins, are indicated. (B) The inhibition of neuromuscular transmission by 20 nM native dichain (■) and 80 nM recombinant dichain (□) was assessed using the mouse hemi-diaphragm preparation. The left phrenic nerve was stimulated at 37°C and the muscle contraction and paralysis times recorded as before (42). Values are the means ( $\pm$  SD) obtained from three different experiments; the 100% value represents the initial muscle contraction evoked by supramaximal stimulation.



4°C with 1 nM TeTx for an additional 30 min. Compared to the control, the times taken for TeTx-induced paralysis were extended by each of the chain preparations but not by MBP (Table I); this demonstrated that the HC and H<sub>c</sub> produced in *E. coli* possess the ability to bind to the ecto-acceptors. However, native HC was about 2-fold more potent than its recombinant counterpart in inhibiting nerve-evoked muscle tension; also, the potency of r-H<sub>c</sub> was half that of r-HC (Table I). Not surprisingly, the native HC is the most efficient antagonist because of it being produced by *Clostridium tetani* as a single-chain protein that includes LC; this may help to induce a conformation of HC that is optimal for binding to the neuronal ecto-acceptors. In contrast, r-HC and r-H<sub>c</sub> were produced in *E. coli* as a fusion products and had to be cleaved with proteases before being subjected to purification. To explore whether these various binding fragments could displace the bound toxin, a series of related experiments was carried out. After the diaphragms were first incubated in mKR buffer with 1 nM TeTx at 4°C for 30 min, to allow TeTx to become bound to the neuronal membrane ecto-acceptors, HC, r-HC, or r-H<sub>c</sub>, were added and further incubated in the same buffer for 60 min at 4°C. The tissue samples were then washed with mKR and KR before the temperature was raised to 37°C and paralysis monitored, as before. The control diaphragms, incubated with 1 nM TeTx only but otherwise processed similarly, became paralysed in  $240 \pm 10$  min; however, the paralysis times for the experimental groups were not significantly different, demonstrating that neither H<sub>c</sub> nor HC could displace the bound toxin.

**Reconstitution of Recombinant Dichain and Assessment of Its Biological Activity In Vivo and In Vitro**—To ascertain whether HC and LC made in *E. coli* can associate and form an interchain disulphide bridge, and to establish if the resultant dichain molecule retains its biological activity, equimolar amounts were mixed and dialysed against re-naturing buffer. SDS-PAGE under non-reducing conditions revealed that reconstitution of the recombinant chains occurred but to a somewhat lesser extent than that observed for their counterparts isolated from TeTx (Fig. 5A; Table II). The biological activity of recombinant dichain was assessed *in vitro* on a mouse nerve-muscle preparation;

80 nM of dichain generated by the two recombinant chains caused neuromuscular paralysis within  $150 \pm 10$  min (Fig. 5B), representing 25% of the neuromuscular activity exhibited by dichain formed with the individual chains isolated from natural TeTx. This is a very satisfactory level of activity for totally recombinant toxin particularly considering that the dichain made using recombinant LC and natural HC possesses lower activity than that reconstituted using both of the chains isolated from TeTx (33, 34). When the recombinant dichain was tested *in vivo*, it displayed appreciable lethality in mice (Table II), its toxicity being 300 ng/kg; thus, it is 40-fold less toxic in the whole animal than the dichain made from the natural chains, compared with only the 4-fold difference found using the neuromuscular paralysis assay. Most importantly, when sub-lethal doses of the recombinant dichain were injected in the neck region of mice, a spastic paralysis was induced; the symptoms produced were indistinguishable from those caused by native TeTx that results from blockade of transmitter release at inhibitory synapses in the spinal cord. However, when r-HC (10 µg) and r-LC (10 µg) were mixed together and immediately injected into mice, the recombinant chain mixture failed to produce typical tetanus symptoms *in vivo* (Table II), in keeping with the requirement for an intact disulphide bond between the LC and the HC. Thus, the dichain produced using both HC and LC synthesised in *E. coli* can be refolded in a conformation that is capable of ecto-acceptor binding, internalisation, axonal retrograde transport and blockade of the release of inhibitory transmitters in spinal cord inter-neurons.

## DISCUSSION

The successful high level expression of r-H<sub>c</sub> and r-HC, for the first time, allowed examination of their effects on TeTx-induced neuromuscular paralysis; also, the recombinant technology established herein offers the important advantage of avoiding the difficulty of removing trace amounts of contaminating LC or dichain from HC or H<sub>c</sub>, as normally isolated from the native toxin. The r-HC and r-H<sub>c</sub> preparations were non-toxic but, like their native counterparts (13, 14), antagonised the neuromuscular activity of TeTx by competing for binding to ecto-acceptors and, thus, delaying the subsequent internalisation and intracellular action of TeTx. Neither native HC, r-HC, nor r-H<sub>c</sub> was capable of slowing neuromuscular paralysis after TeTx had first bound to the target cells; this is indicative of very tight binding of the native toxin, though the precise nature of the ecto-acceptor still remains unclear. It has been proposed that gangliosides can act as receptors for TeTx (15, 46) but proteins are also thought to be involved in its neuronal binding (47, 48). Such formation of toxin complexes with ganglioside and protein receptor(s) has been suggested to underlie the specific high-affinity binding that culminates in internalisation and translocation (49). The twofold higher potency of native HC relative to r-HC accords with the fact that native HC is produced and folded, under optimal conditions, as a single chain; later, it is cleaved to yield the dichain toxin. The lower binding efficiency of r-HC could arise from less perfect folding because of being produced in *E. coli* as a fusion and exposed to enzymatic cleavage and a lengthy purification process; however, there was no significant difference between the CD spectra of r-HC and native

TABLE II. Mouse lethality of TeTx and dichains reconstituted from either native or recombinant chains.

Samples <sup>a</sup>	Mouse lethality <sup>b</sup> (LD <sub>50</sub> : ng/kg body wt)	Covalently linked dichain (%)
TeTx	1	—
Dichains reconstituted using:		
Native chains (HC & LC)	7.5	62 <sup>c</sup>
Recombinant chains (HC & LC)	300	44 <sup>c</sup>
r-HC and r-LC mixed but not reconstituted <sup>d</sup>	> 10 <sup>e</sup>	—

<sup>a</sup>Equimolar amount of native or recombinant HC and LC were mixed and dialysed against 50 mM Tris-HCl (pH 8.4), 1 M NaCl, 2 M urea, and 20 mM DTT with stirring at 4°C for 16 h, followed by dialysis against 50 mM Tris-HCl (pH 8.4) and 600 mM glycine without agitation at 4°C for 72 h. <sup>b</sup>Mouse toxicity was defined as that amount of toxin (ng/kg) which kills 50% of injected mice within four days, as detailed previously (41). <sup>c</sup>The formation of dichain was monitored by SDS-PAGE in the absence of reducing reagent (Fig. 5) and the content of dichain species quantified by densitometric scanning of Coomassie stained gels. <sup>d</sup>Recombinant chains [HC (10 µg) and LC (10 µg)] which were mixed together just before being injected into mice. Mean values are shown from at least three different experiments.

HC indicating that their gross folded structures are similar, consistent with the acceptable level of biological activity observed for the recombinant material. Notably, r-H<sub>C</sub> proved less effective than r-HC in retarding the neuromuscular paralysis caused by TeTx, possibly because the latter adopts a more native conformation due to the presence of the H<sub>N</sub> fragment and/or the H<sub>N</sub> may contribute to the ecto-acceptor binding. Although the native HC, r-HC, and r-H<sub>C</sub> delayed the onset of paralysis by TeTx of neuromuscular junction, the antagonism exhibited was not complete even when a vast molar excess of each over TeTx was used. This might be explained by the fact that these suboptimally folded fragments could not saturate all the TeTx acceptors on the nerve terminals; therefore, the highly potent TeTx may still interact with the acceptors via a very limited number of unoccupied sites and subsequently block transmitter release.

It was also demonstrated, for the first time, that r-HC can be successfully reconstituted with expressed LC, forming a disulphide-bridged dichain. The resultant complete recombinant toxin proved to be only 4-fold less potent than the two reconstituted native chains. However, when tested *in vivo*, the recombinant dichain proved 40-fold less toxic than its native equivalent; this may be indicative of a lower amount of recombinant protein being efficiently transported into CNS, the main target for TeTx. Less perfect folding of the recombinant chains themselves may account for the lower percentage of dichain formation, and this may also increase its susceptibility to degradation *in vivo*, resulting in the reduced lethality observed. In contrast, free r-LC and r-HC when briefly mixed together before injection did not exhibit any toxicity in mice, even when large quantities of each were employed. This clearly indicates that an intact interchain disulphide bridge between LC and HC is required, a bond which cannot form simply by mixing the two separate chains together for a short time. This emphasises the importance of the disulphide bridge in the membrane translocation of TeTx LC into the cytosol of targetted neurons, a result previously observed (50).

The novelty of this study is that functional dichain has been produced from r-HC and r-LC, expressed separately in *E. coli*. A CNS-directed drug-delivery vehicle based upon a recombinant dichain, formed from r-HC and an enzymatically-inactivated isoform of r-LC (33), is now feasible. Additionally, the expressed non-toxic whole protein may produce better immunological protection against tetanus than chemically-inactivated TeTx. Indeed, the innocuous dichain toxin gave a higher antibody titre than that of r-H<sub>C</sub> alone (28); also, antibodies raised against the LC were found to give protection against challenge with TeTx (28, 51). Thus, non-toxic mutants of TeTx have the potential to provide improved vaccines for tetanus which kills millions of people yearly; these would be safer to handle, easier to produce and could be purified by affinity rather than conventional chromatography. Lastly, the proven feasibility of producing functional r-HC and r-H<sub>C</sub> provides scope for investigating the HC and H<sub>C</sub> by site-directed mutagenesis to elucidate their roles in binding, internalisation and retrograde transport of TeTx.

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## REFERENCES

- Wellhöner, H.H. (1992) Tetanus and botulinum neurotoxins in *Handbook of Experimental Pharmacology* (Herken, H. and Hucho, F. eds.), Vol 102, pp. 357-417, 'Selective Neurotoxicity,' Chapter 11, Springer-Verlag, Berlin
- Weller, U., Dauzenroth, M.-E., Meyer zu Heringdorf, D., and Habermann, E. (1989) Chains and fragments of tetanus toxin: Separation, reassociation and pharmacological properties. *Eur. J. Biochem.* 182, 649-656
- Simpson, L.L. (1981) The origin, structure and pharmacological activity of botulinum toxin. *Rev. Pharmacol.* 33, 155-188
- Simpson, L.L. (1989) Peripheral actions of the botulinum toxins in *Botulinum Neurotoxin and Tetanus Toxin* (Simpson, L.L., ed.) pp. 153-178, Academic Press, New York
- Dolly, J.O. (1992) *Handbook of Experimental Pharmacology* (Herken, H. and Hucho, F., eds.) Vol. 102, pp. 681-717, Springer-Verlag, Berlin
- Habermann, E., Bigalke, H., Dreyer, F., and Streitzig, P. (1980) *Natural Toxins* (Eaker, D. and Wadström, T., eds.), pp. 593-599, Pergamon Press, Oxford
- Niemann, H. (1991) Molecular biology of clostridial neurotoxins in *Sourcebook of Bacterial Protein Toxins*, 303-348
- Schiavo, G., Benfenati, F., Poulain, B., Rossetto, O., Delaureto, P.P., Dasgupta, B.R., and Montecucco, C. (1992) Tetanus and botulinum-B neurotoxins block neurotransmitter release by proteolytic cleavage of synaptobrevin. *Nature* 359, 832-835
- Link, E., Edelmann, L., Chou, J.H., Binz, T., Yamasaki, S., Eisel, U., Baumert, M., Südhof, T.C., Niemann, H., and Jahn, R. (1992) Tetanus toxin action—inhibition of neurotransmitter release linked to synaptobrevin proteolysis. *Biochem. Biophys. Res. Commun.* 189, 1017-1023
- Facchiano, F. and Luini, A. (1992) Tetanus toxin potentially stimulates tissue transglutaminase—a possible mechanism of neurotoxicity. *J. Biol. Chem.* 267, 13267-13271
- Facchiano, F., Benfenati, F., Valtorta, F., and Luini, A. (1993) Covalent modification of synapsin-I by a tetanus toxin-activated transglutaminase. *J. Biol. Chem.* 268, 4588-4591
- Helting, T.B. and Zwisler, O. (1977) Structure of tetanus toxin. I. Breakdown of the toxin molecule and discrimination between polypeptide fragments. *J. Biol. Chem.* 252, 187-193
- Simpson, L.L. (1984) Fragment C of tetanus toxin antagonizes the neuromuscular blocking properties of native tetanus toxin. *J. Pharmacol. Expt. Ther.* 228, 600-604
- Simpson, L.L. (1984) The binding fragment from tetanus toxin antagonises the neuromuscular blocking actions of botulinum toxin. *J. Pharmacol. Expt. Ther.* 229, 182-187
- Morris, N.P., Consiglio, E., Kohn, L.D., Habig, W.H., Hardegree, M.C., and Helting, T.B. (1980) Interaction of fragments B and C of tetanus toxin with neural and thyroid membranes and with gangliosides. *J. Biol. Chem.* 255, 6071-6076
- Goldberg, R.L., Costa, T., Habig, W.H., Kohn, L.D., and Hardegree, M.C. (1981) Characterization of fragment C and tetanus toxin binding to rat brain membranes. *Mol. Pharmacol.* 20, 565-570
- Weller, U., Taylor, C.F., and Habermann, E. (1986) Quantitative comparison between tetanus toxin, some fragments and toxoid for binding and axonal transport in the rat. *Toxicon* 24, 1055-1063
- Boquet, P. and Duflot, E. (1982) Tetanus toxin fragment forms channels in lipid vesicles at low pH. *Proc. Natl. Acad. Sci. USA* 79, 7614-7618
- Hogy, B., Dauzenroth, M.E., Hudel, M., Weller, U., and Habermann, E. (1992) Increase of permeability of synaptosomes and liposomes by the heavy chain of tetanus toxin. *Toxicon* 30, 63-76
- Ashton, A.C., Li, Y., Doussau, F., Weller, U., Dougan, G., Poulain, B., and Dolly, J.O. (1995) Tetanus toxin inhibits neuroexocytosis even when its Zn<sup>2+</sup>-dependent protease activity



- is removed. *J. Biol. Chem.* 270, 31386-31390
21. Penner, R., Neher, E., and Dreyer, F. (1986) Intracellularly, injected tetanus toxin inhibits exocytosis in bovine adrenal chromaffin cells. *Nature* 324, 76-78
  22. Fairweather, N.F. and Lyness, V.A. (1986) The complete nucleotide-sequence of tetanus toxin. *Nucleic Acids Res.* 14, 7809-7812
  23. Fairweather, N.F., Lyness, V.A., Pickard, D.J., Allen, G.F., and Thomson, R. (1986) Cloning, nucleotide sequencing and expression of tetanus toxin fragment C in *Escherichia coli*. *J. Bacteriol.* 165, 21-27
  24. Romanos, M.A., Makoff, A.J., Fairweather, N.F., Beesley, K.M., Slater, D.E., Rayment, F.B., Payne, M.M., and Clare, J.J. (1991) Expression of tetanus toxin fragment-C in yeast—gene synthesis is required to eliminate fortuitous polyadenylation sites in AT-rich DNA. *Nucleic Acids Res.* 19, 1461-1467
  25. Charles, I.G., Rodgers, B.C., Makoff, A.J., Chatfield, S.N., Slater, D.E., and Fairweather, N.F. (1991) Synthesis of tetanus toxin fragment-C in insect cells by use of a baculovirus expression system. *Inf. Immun.* 59, 1627-1632
  26. Halpern, J.L., Habig, W.H., Neale, E.A., and Stibitz, S. (1990) Cloning and expression of functional fragment C of tetanus toxin. *Inf. Immun.* 58, 1004-1009
  27. Halpern, J.L. and Loftus, A. (1993) Characterization of the receptor-binding domain of tetanus toxin. *J. Biol. Chem.* 268, 11188-11192
  28. Figueiredo, D., Turcotte, C., Frankel, G., Li, Y., Dolly, O., Wilkin, G., Marriott, D., Fairweather, N., and Dougan, G. (1995) Characterization of recombinant tetanus toxin derivatives suitable for vaccine development. *Inf. Immun.* 63, 3218-3221
  29. Schwab, M.E., Suda, K., and Thoenen, H. (1979) Selective retrogradetrans-synaptic transfer of a protein, tetanus toxin, subsequent to its retrograde axonal transport. *J. Cell Biol.* 82, 798-810
  30. Fishman, P.S. and Savitt, J.M. (1989) Transsynaptic transfer of retrogradely transported tetanus protein peroxidase conjugates. *Exp. Neurol.* 106, 197-203
  31. Fishman, P.S., Savitt, J.M., and Farrand, D.A. (1990) Enhanced CNS uptake of systemically administered proteins through conjugation with tetanus C-fragment. *J. Neurol. Sci.* 98, 311-325
  32. Dqbrenis, K., Joseph, A., and Rattazzi, M.C. (1992) Neuronal lysosomal enzyme replacement using fragment-C of tetanus toxin. *Proc. Natl. Acad. Sci. USA* 89, 2297-2301
  33. Li, Y., Foran, P., Fairweather, N.F., de Paiva, A., Weller, U., Dougan, G., and Dolly, J.O. (1994) A single mutation in the recombinant light chain of tetanus toxin abolishes its proteolytic activity and removes the toxicity seen after reconstitution with native heavy chain. *Biochemistry* 33, 7014-7020
  34. Fairweather, N.F., Sanders, D., Slater, D., Hudel, M., Habermann, E., and Weller, U. (1993) Production of biologically-active light-chain of tetanus toxin in *Escherichia coli*—evidence for the importance of the C-terminal 16 amino-acids for full biological-activity. *FEBS Lett.* 323, 218-222
  35. Zhou, L.Q., de Paiva, A., Liu, D., Aoki, R., and Dolly, J.O. (1995) Expression and purification of the light chain of botulinum neurotoxin A: a single mutation abolishes its cleavage of SNAP-25 and neurotoxicity after reconstitution with the heavy chain. *Biochemistry* 34, 15175-15181
  36. Hermanson, G.T., Mallia, A.K., and Smith, P.K. (1992) Immobilised affinity ligand techniques, Academic Press, San Diego, California
  37. Tous, G.I., Fausnaugh, J.L., Akinyosoye, O., Lackland, H., Winter-Cash, P., Vitorica, F.J., and Stein, S. (1989) Amino acid analysis on polyvinylidene difluoride membranes. *Anal. Biochem.* 179, 50-55
  38. Eisel, U., Jarausch, W., Goretzki, K., Henschen, A., Engels, J., Weller, U., Hudel, M., Habermann, E., and Niemann, H. (1986) Tetanus toxin: primary structure, expression in *E. coli*, and homology with botulinum toxins. *EMBO J.* 5, 2495-2502
  39. Mao, D. and Wallace, B.A. (1984) Differential light scattering and absorption flattening optical effects are minimal in the circular dichroism spectra of small unilamellar vesicles. *Biochemistry* 23, 2667-2673
  40. Chang, C.T., Wu, C.S.C., and Yang, J.T. (1978) Circular dichroic analysis of protein conformation: inclusion of the b-turns. *Anal. Biochem.* 91, 13-31
  41. Maisey, E.A., Wadsworth, J.D.F., Poulain, B., Shone, C.C., Melling, J., Gibbs, P., Tauc, L., and Dolly, J.O. (1988) Involvement of the constituent chains of botulinum neurotoxin A and B in the blockade of neurotransmitter release. *Eur. J. Biochem.* 177, 683-691
  42. de Paiva, A., Poulain, B., Lawrence, G.W., Shone, C.C., Tauc, L., and Dolly, J.O. (1993) A role for the interchain disulfide or its participating thiols in the internalization of botulinum neurotoxin-A revealed by a toxin derivative that binds to ecto-acceptors and inhibits transmitter release intracellularly. *J. Biol. Chem.* 268, 20838-20844
  43. de Paiva, A. and Dolly, J.O. (1990) Light chain of botulinum neurotoxin is active in mammalian motor nerve terminals when delivered via liposomes. *FEBS Lett.* 277, 171-174
  44. Robinson, J.P. and Hash, J.H. (1982) A review of the molecular structure of tetanus toxin. *Mol. Cell Biochem.* 48, 33-44
  45. Defilippis, V., Vangelista, L., Schiavo, G., Tonello, F., and Montecucco, C. (1995) Structural studies on the zinc-endopeptidase light chain of tetanus neurotoxin. *Eur. J. Biochem.* 229, 61-69
  46. van Heyningen, W.E. (1974) Gangliosides as membrane receptors for tetanus toxin, cholera toxin and serotonin. *Nature* 249, 415-417
  47. Bigalke, H., Heller, L., Bizzini, B., and Habermann, E. (1981) Tetanus toxin and botulinum A toxin inhibit release and uptake of various transmitters, as studied with particulate preparations from rat brain and spinal cord. *Naunyn-Schm. Arch. Pharmacol.* 316, 244-251
  48. Lazarovici, P. and Yavin, E. (1986) Affinity-purified tetanus neurotoxin interaction with synaptic membranes: properties of a protease-sensitive receptor component. *Biochemistry* 25, 7047-7054
  49. Montecucco, C. (1986) How do tetanus and botulinum toxins bind to neuronal membranes? *TiBS* 11, 314-317
  50. Schiavo, G., Papini, E., Genna, G., and Montecucco, C. (1990) An intact interchain disulfide bond is required for the neurotoxicity of tetanus toxin. *Inf. Immun.* 58, 4136-4141
  51. Lin, C.S., Habig, W.H., and Hardegree, M.C. (1985) Antibodies against the light chain of tetanus toxin in human sera. *Inf. Immun.* 49, 111-115